

Zinc Is Not Required for Activity of TPO Agonists Acting at the c-Mpl Receptor Transmembrane Domain

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uman thrombopoietin (TPO) is a circulatory cytokine that is the primary regulator of megakarocytopoiesis and platelet production. In addition, TPO is an early acting synergistic cytokine regulating quiescence and proliferation of hematopoietic stem cells (1, 2). TPO binds to its receptor, human myeloproliferative leukemia virus oncogene (c-Mpl), and initiates an intracellular signaling cascade (1), resulting in production of platelets essential in wound healing (3, 4). Although TPO alone has been used clinically, concerns about its immunogenicity and, to some extent, cost of the native cytokine and conjugates have limited widespread clinical application (5, 6). The development of small, synthetic, orally active agonist compounds of TPO would be very beneficial to treat thrombocytopenia and to drive hematopoietic stem cells to desired cellular fates (7).

Random screening has identified a relatively diverse range of synthetic compounds that act as c-Mpl agonists, including pyrazol-4-ylidenehydrazines (*8*), salicylaldehyde thiosemicarbazones (*9*), naphtho(1,2-*d*) imidazoles (*10*), xanthocillins (*11*), benzimidazoles (*12*), hydrazinothiophenes (*13*), arylthiazoles (*14*), benzamides (*15–18*), benzocarbazoles (*19, 20*), and butyzamides (*21*). One of these classes has yielded a new drug, Eltrombopag, which has recently received registration from the FDA for the treatment of thrombocytopenia (*20*).

Unusually, these compounds interact with the transmembrane helix of the receptor. In the human and chimpanzee receptor, the transmembrane helix contains a unique histidine residue that, *via* single point knock out and knock in mutation experiments, has been shown to be essential for the activity of this class of TPO agonist (*15*). No other animal species carry this histidine mutation, and their c-Mpl receptors are unaffected by these agonists (*13, 15, 22*).

Early work on this class of mimetic (10) identified a molecular feature that was hypothesized to chelate zinc ions and carry them into the c-Mpl transmembrane domain, where they interact with H499 to activate the receptor (see Figure 1).

Subsequent work attributed this "zincchelating" role to the 15 or so chemotypes that exhibit TPO agonist activity *via* this unusual mechanism (*19, 23*). Evidence for this hypothesis was published in a patent by Delorme *et al.* (*24*). They reported experiments in which two pyrazol-4-ylidenehydrazines exhibited activity against mutant C610H and double mutant G607T murine GCSF receptor constructs in the presence of added zinc ions. Chelation of zinc by EDTA abolished receptor activity. This single experiment using a receptor construct was extrapolated to **ABSTRACT** Molecules that mimic the cytokine thrombopoietin that act by an atypical mechanism of binding to a receptor transmembrane (TM) domain are widely understood to require zinc for their biological activity. We investigated potent thrombopoietin mimetics from three chemical classes including the recently registered drug Eltrombopag, which operate via this novel mechanism, to determine whether zinc is essential for inducing cell proliferation. Using addition of zinc and a potent metal chelator, we show that the existing paradigm is incorrect and the compounds exhibit excellent thrombopoietin-mimetic activity even in the presence of high concentrations of EDTA. The implications of these findings for the mechanism of action are discussed.

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Figure 1. Comparison of pyrazol-4-ylidenehydrazine TPO mimic A and thiosemicarbazone TPO mimic B, illustrating the three key features for potent thrombopoietic activity: central metal chelate, lipophilic end portion, and a distal acidic functionality (from ref 9, used with permission).

conclude that all compounds of this type required zinc to function as TPO agonists. No primary, peer-reviewed publication has formally demonstrated this general prerequisite for zinc, although subsequent NMR and mutation experiments on chimeric constructs of c-Mpl (*25*) cast doubt on the hypothesis. Our preliminary work caused us also to question the role of zinc in compounds acting at the TM domain of c-Mpl. In particular, the high TPO agonist activity of the benzocarbazoles (*19, 20*) that are covalently constrained and lack a strong chelating group is inconsistent with a requirement for zinc.

We screened four mimetics of TPO using a human transfected factor-dependent murine hematopoietic cell line (FD-MpI) in combination with a high concentration of zinc sulfate or EDTA. We wished to determine whether high or enhanced levels of zinc could improve the binding of these mimetics to the c-MpI receptor and enhance cell proliferation and whether high levels of EDTA eliminate mimetic activity.

We assessed the effect of zinc and EDTA across a range of concentrations on the growth and viability of FD-Mpl cells growing in media supplemented with rhTPO (30 ng mL⁻¹) to ensure that the metal ion and the chelating agent did not affect cell proliferation on their own. We saw no statistically

significant difference between the numbers of cells cultured with rhTPO only, rhTPO plus 100 µM zinc, and rhTPO plus 100 µM EDTA.

The TPO- and TPO+ controls all gave the expected results (see Methods), with either no viable cells or strong growth at 30 ng mL⁻¹ of rhTPO respectively. Figure 2 shows cell proliferation results for compounds **1**, **2a**, **2b**, and **3** with and without

added Zn²⁺ and with added EDTA. Cell proliferation was normalized using the cell numbers obtained from culture in media supplemented with 30 ng mL⁻¹ (0.84 nM) rhTPO expressed as a percentage. Asterisks indicate where the responses were different at the 95% confidence limit.

The effect of added high dose zinc and chelator on factor-dependent cells stimulated by Eltrombopag (1) is summarized in Figure 2, panel a. Added ZnSO₄ had negligible effect on cell growth (cell numbers comparable to those with 10 and 30 ng mL⁻¹ rhTPO). Addition of EDTA to 1 gave slightly enhanced cell numbers and more variable results, but the difference was not statistically significant. The addition of zinc and EDTA appear to have no effect, giving cell numbers virtually identical to 1 alone at 1 μ M. The ED₅₀ values for the control and both treatment groups were essentially the same.

Figure 2, panels b and c summarize the results of the experiments adding Zn or EDTA to the pyrazolohydrazinonaphthalene (PHN) compounds **2a,b**. The compounds gave a good dose—response curve and high cell numbers when used alone, although marginally less than **1**. The addition of Zn-SO₄ to **2a** gave cell responses identical to those with **2a** alone (Figure 2, panel b). The addition of EDTA did not significantly

change cell growth compared that of the control (**2a** alone), generating a dose—response curve very similar to that of **2a** alone, and **2a** with added Zn. The graph shows that the ED_{50} values for all three samples were essentially the same within experimental error, as was observed for **1**

The addition of ZnSO₄ to **2b** marginally decreased cell numbers at 1 and 10 μ M compared to the control of compound **2b** alone and also produced an EC₅₀ value slightly higher than that of **2b** alone. The addition of EDTA marginally decreased cell growth below that of the control but exhibited an EC₅₀ similar to that of the control (Figure 2, panel c). Dose response curves for the three conditions were similar, but the maximum proliferation for the treated groups was lower than the control.

The benzocarbazole, 3, was an important compound because it has a conformationally constrained structure and no clear metal chelating group, suggesting it does interact with zinc strongly. However, it clearly exhibited a different response compared to the other agonists (Figure 2, panel d), although the differences were still small. Both treated groups showed reduced maximum proliferation compared to the control, as was seen with other agonists. EDTA-treated cells had a dose-response curve and EC50 value similar to those of the control. Interestingly, the zinc-treated cells exhibited a EC_{50} value lower than those of the control and EDTA-treated cells, and this difference was statistically significant. At the highest dose tested, there were crystals in all cultures, indicating precipitation. Paradoxically, the class of compounds least likely to chelate zinc showed a small decrease in EC₅₀ on addition of zinc ions.

Thrombopoietin is a critical cytokine for generating megakaryocytes and platelets from hematopoietic stem and progenitor cells cultured *ex vivo*. Small molecule mimics that interact with the TM domain of c-Mpl may be replacements for native TPO. It is im-

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Figure 2. a) Growth-response curves of factor-dependent cells with Eltrombopag 1 alone, with Eltrombopag 1 and 100 μ M ZnSO₄, and with Eltrombopag 1 and 100 μ M EDTA. b) Growth-response curves of factor-dependent cells with PHN 2a alone, with 2a and 100 μ M ZnSO₄, and with 2a and 100 μ M EDTA (note that the black curve lies directly under the green curve). c) Growth-response curves of factor-dependent cells with PHN 2b alone, with 2b and 100 μ M ZnSO₄, and with 2b and 100 μ M EDTA. d) Growth-response curves of factor-dependent cells with PHN 2b alone, with 2b and 100 μ M ZnSO₄, and with 2b and 100 μ M EDTA. d) Growth-response curves of factor-dependent cells with benzocarbazole 3 alone, with 3 and 100 μ M ZnSO₄, and with 3 and 100 μ M EDTA.

portant to understand how these molecules work so that culture conditions be deployed that ensure their maximal bioactivity. As previous reports had suggested that zinc was essential for bioactivity (*10, 19, 23*), it is also important to consider the baseline levels of zinc normally present in most culture media like DMEM, supplemented with 10% FCS. This has been estimated to be 4 μ M (*26*). *In vivo*, zinc is bound to albumin in the circulation, where it is maintained at approximately 1 μ g mL⁻¹ in mammals. The total intracellular zinc content of a typical fibroblast, in cell culture, is 0.25 fmol or \sim 200 μ M (*29*). These levels are similar to the concentrations of the TPO agonists used

in our experiments and would be sufficient to activate them if zinc binding was necessary for their activities.

EDTA binds zinc extremely strongly with a K_d of $\sim 10^{-16}$, compared to a K_d for magnesium of 10^{-9} and K_d for calcium of 10^{-11} , and will selectively deplete a solution of free zinc ions in the presence of other free diva-

lent ions, even if they are present at high concentration (28). The kinetic behavior of zinc is summarized by Nyborg (29). Intraventricular injection of EDTA has been reported to deplete intracellular zinc levels so that the levels of EDTA used for our experiments should remove any incidental zinc in the culture medium and in the cells (30). Accordingly, our data strongly suggest that TM acting mimics do not require zinc for their activity.

We also conducted a computational study of structure—activity relationships in ~400 members of this class of TPO agonists (*31*). The model was very successful in explaining the molecular features required for agonist activity and did not require zinc chelation. Our study suggested that the putative metal binding moiety in most agonists has a different role than binding zinc. The heteroatoms in the linker region contribute to stabilization of a pseudo planar conformation of the agonists *via* intramolecular hydrogen bonds, amide planarity, tautomeric effects, and π conjugation, suggesting this conformation is essential for activity.

Dimerization of c-Mpl and signaling by agonists will be dependent on intramolecular distances between ligands and receptors and the fit of the two surfaces which, from our results, does not require the presence of a zinc ion. The exact molecular mechanism by which the small molecule agonist acting at or near the key histidine residue in the transmembrane helix is far from clear.

It is also not clear why the highest levels of zinc used in this study sometimes enhance the cell proliferation activity of the small molecule agonists. The levels used are much higher than physiological, so the observations may not be biologically relevant. These high levels of zinc did not significantly alter the activity of TPO, and not all classes of small molecule TPO agonists showed substantial effects. We speculate that the enhancement may be due to additional stabilization of the required binding conformation of agonist by zinc chelation in the linker region (*31*). Our QSAR studies suggest that agonists wrap around the transmembrane helix and that the linker region heteroatoms interact with transmembrane histidines. This could lock them into a position where the Lewis base region of agonists interact with the extracellular juxtamembrane domain of c-Mpl and cause receptor dimerization.

In conclusion, our experiments with a TPO-dependent cell line show that zinc is not required for agonist activity of four compounds in three chemical classes, including the recently registered drug Eltrombopag, that act *via* the atypical transmembrane mechanism. While the linker region of TPO agonist compounds is still able to chelate zinc in some cases, this is not essential for activity. The previous mechanistic paradigm needs revising, as the addition of EDTA does not abolish activity.

METHODS

Compounds **1**, **2a**, and **2b** (Supplementary Figure 1) were prepared by published methods (*8*, *24*). The benzocarbazole **3** was prepared by the method of Marsilje *et al.* (*19*, *32*) and references cited therein. The identities of the four synthesized compounds were confirmed by comparison of their ¹H NMR spectra with those reported in the literature (see Supporting Information).

Biological Screening. A TPO-dependent cell line was created by retroviral transduction of the murine hematopoietic cell line FDCP-1 (*33*) with the ecotropic Psi2 vector containing full-length human c-MPL. These cells are absolutely dependent on human TPO for survival and were used throughout the study for bioassay of all c-Mpl agonists. The FD-Mpl line was validated with native ligand TPO and Eltrombopag, a potent small molecule mimetic of TPO acting *via* the transmembrane mechanism. The dose—response rhTPO curve is shown in Supplementary Figure 2.

Proliferation of 5000 FD-Mpl cells/96 well in the absence of added Zn²⁺, 100 mM Zn²⁺ as ZnSO₄-7H₂O (BDH, Poole, England), and 100 mM EDTA (BDH, Poole, England) was carried out for 48 h at 37 °C (humidified air, 5% CO₂) in DMEM with 10% FCS. After 48 h cell proliferation was measured using a CellTiter-Glo Luminescent Cell Viability Assay (Promega, no. G7570) and expressed as a percentage of cell proliferation obtained in media supplemented with either 30 ng mL⁻¹ of rhTPO, 30 ng mL⁻¹ of rhTPO and 100 μ M EDTA. Importantly, in the absence of activation on the c-Mpl receptor, the FD-Mpl cells die within 48 h (Supplementary Figure 3). We also varied the sequence of addition of reagents, but this produced no change to the outcomes reported here. Statistical analysis was carried out using a two-way ANOVA with interactions comparing factor 1 (control, no added Zn²⁺) with factor 2 (100 μ M added Zn²⁺ or 100 μ M added EDTA).

Supporting Information Available: Structures and characterization of TPO agonist compounds, response of factor-dependent (fd-Mpl) cells to TPO, fd-Mpl cell viability in response to TPO starvation. This material is available free of charge via the Internet at http://pubs.acs.org.

REFERENCES

- 1. Bolam, S. (1997) Thrombopoietin, *Transfus. Sci. 18*, 129–137.
- 2. Kaushansky, K. (1995) Thrombopoietin—basic biology, clinical promise, *Int. J. Hematol.* 62, 7–15.
- Wendling, F. (1999) Thrombopoietin: its role from early hematopoiesis to platelet production, *Haematologica* 84, 158–166.
- Katayama, N., Itoh, R., Kato, T., Sugawara, T., Mahmud, N., Ohishi, K., Masuya, M., Aoki, M., Minami, N., Miyazaki, H., and Shiku, H. (1997) Role for c-Mpl and its ligand thrombopoietin in early hematopoiesis, *Leukemia Lymphoma 28*, 51–56.
- Tiu, R. V., and Sekeres, M. A. (2008) The role of AMG-531 in the treatment of thrombocytopenia in idiopathic thrombocytopenic purpura and myelodysplastic syndromes, *Expert Opin. Biol. Ther.* 8, 1021–1030.
- Haznedaroglu, I. C., Goker, H., Turgut, M., Buyukasik, Y., and Benekli, M. (2002) Thrombopoietin as a drug: biologic expectations, clinical realities, and future directions, *Clin. Appl. Thromb./Hemostasis 8*, 193–212.
- Peeters, K., Stassen, J. M., Collen, D., Van Geet, C., and Freson, K. (2008) Emerging treatments for thrombocytopenia: increasing platelet production, *Drug Discovery Today* 13, 798–806.
- Duffy, K. J., Darcy, M. G., Delorme, E., Dillon, S. B., Eppley, D. F., Erickson-Miller, C., Giampa, L., Hopson, C. B., Huang, Y. F., Keenan, R. M., Lamb, P., Leong, L., Liu, N. N., Miller, S. G., Price, A. T., Rosen, J., Shah, R., Shaw, T. N., Smith, H., Stark, K. C., Tian, S. S., Tyree, C., Wiggall, K. J., Zhang, L., and Luengo, J. I. (2001) Hydrazinonaphthalene and azonaphthalene thrombopoietin mimics are nonpeptidyl promoters of megakaryocytopoiesis, *J. Med. Chem.* 44, 3730–3745.
- Duffy, K. J., Shaw, A. N., Delorme, E., Dillon, S. B., Erickson-Miller, C., Giampa, L., Huang, Y. F., Keenan, R. M., Lamb, P., Liu, N. N., Miller, S. G., Price, A. T., Rosen, J., Smith, H., Wiggall, K. J., Zhang, L. H., and Luengo, J. I. (2002) Identification of a pharmacophore for thrombopoietic activity of small, nonpeptidyl molecules. 1. Discovery and optimization of salicylaldehyde thiosemicarbazone thrombopoietin mimics, J. Med. Chem. 45, 3573–3575.
- Duffy, K. J., Price, A. T., Delorme, E., Dillon, S. B., Duquenne, C., Erickson-Miller, C., Giampa, L., Huang, Y. F., Keenan, R. M., Lamb, P., Liu, N. N., Miller, S. G., Rosen, J., Shaw, A. N., Smith, H., Wiggall, K. J., Zhang, L. H., and Luengo, J. I. (2002) Identification of a pharmacophore for thrombopoietic activity of

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small, non-peptidyl molecules. 2. Rational design of naphtho[1,2-d]imidazole thrombopoietin mimics, *J. Med. Chem.* 45, 3576–3578.

- Sakai, R., Nakamura, T., Nishino, T., Yamamoto, M., Miyamura, A., Miyamoto, H., Ishiwata, N., Komatsu, N., Kamiya, H., and Tsuruzoe, N. (2005) Xanthocillins as thrombopoietin mimetic small molecules, *Bioorg. Med. Chem.* 13, 6388–6393.
- Safonov, I. G., Heerding, D. A., Keenan, R. M., Price, A. T., Erickson-Miller, C. L., Hopson, C. B., Levin, J. L., Lord, K. A., and Tapley, P. M. (2006) New benzimidazoles as thrombopoietin receptor agonists, *Bioorg. Med. Chem. Lett.* 16, 1212–1216.
- Nakamura, T., Miyakawa, Y., Miyamura, A., Yamane, A., Suzuki, H., Ito, M., Ohnishi, Y., Ishiwata, N., Ikeda, Y., and Tsouruzoe, N. (2006) A novel nonpeptidyl human c-Mpl activator stimulates human megakaryopoiesis and thrombopoiesis, *Blood* 107, 4300–4307.
- Kalgutkar, A. S., Driscoll, J., Zhao, S. X., Walker, G. S., Shepard, R. M., Soglia, J. R., Atherton, J., Yu, L., Mutlib, A. E., Munchhof, M. J., Reiter, L. A., Jones, C. S., Doty, J. L., Trevena, K. A., Shaffer, C. L., and Ripp, S. L. (2007) A rational chemical intervention strategy to circumvent bioactivation liabilities associated with a nonpeptidyl thrombopoietin receptor agonist containing a 2-amino-4-arylthiazole motif, *Chem. Res. Toxicol. 20*, 1954–1965.
- Yamane, N., Tanaka, Y., Ohyabu, N., Yamane, S., Maekawa, K., Ishizaki, J., Suzuki, R., Itoh, T., and Takemoto, H. (2008) Characterization of novel nonpeptide thrombopoietin mimetics, their species specificity and the activation mechanism of the thrombopoietin receptor, *Eur. J. Pharmacol.* 586, 44–51.
- 16. Yamane, N., Takahashi, K., Tanaka, Y., Kato, K., Takayama, M., Ohyabu, N., Shiota, T., Takenaka, H., Yoshida, Y., Hara, S., Murashi, T., Nakamura, E., Nishitani, Y., Ishizaki, J., Yamane, S., Nagata, K., Koizumi, K., Yutsudo, T., Suzuki, R., Itoh, T., and Takemoto, H. (2008) Discovery of novel non-peptide thrombopoietin mimetic compounds that induce megakaryocytopoiesis, *Biosci. Rep.* 28, 275–285.
- Reiter, L. A., Subramanyam, C., Mangual, E. J., Jones, C. S., Smeets, M. I., Brissette, W. H., McCurdy, S. P., Lira, P. D., Linde, R. G., Li, Q. F., Zhang, F., Antipas, A. S., Blumberg, L. C., Doty, J. L., Driscoll, J. P., Munchhof, M. J., Ripp, S. L., Shavnya, A., Shepard, R. M., Sperger, D., Thomasco, L. M., Trevena, K. A., Wolf-Gouveia, L. A., and Zhang, L. L. (2007) Pyrimidine benzamide-based thrombopoietin receptor agonists, *Bioorg. Med. Chem. Lett.* 17, 5447–5454.
- Reiter, L. A., Jones, C. S., Brissette, W. H., McCurdy, S. P., Abramov, Y. A., Bordner, J., DiCapua, F. M., Munchhof, M. J., Rescek, D. M., Samardjiev, I. J., and Withka, J. M. (2008) Molecular features crucial to the activity of pyrimidine benzamide-based thrombopoietin receptor agonists, *Bioorg. Med. Chem. Lett.* 18, 3000–3006.
- Marsilje, T. H., Alper, P. B., Lu, W. S., Mutnick, D., Michellys, P. Y., He, Y., Karanewsky, D. S., Chow, D., Gerken, A., Lao, J. M., Kim, M. J., Seidel, H. M., and Tian, S. S. (2008) Optimization of small molecule agonists of the thrombopoietin (Tpo) receptor derived from a benzo[a]carbazole hit scaffold, *Bioorg. Med. Chem. Lett.* 18, 5259–5262.

- Alper, P. B., Marsilje, T. H., Mutnick, D., Lu, W. S., Chatterjee, A., Roberts, M. J., He, Y., Karanewsky, D. S., Chow, D., Lao, J. M., Gerken, A., Tuntland, T., Liu, B., Chang, J., Gordon, P., Seidel, H. M., and Tian, S. S. (2008) Discovery and biological evaluation of benzo[a]carbazole-based small molecule agonists of the thrombopoietin (Tpo) receptor, *Bioorg. Med. Chem. Lett.* 18, 5255–5258.
- Nogami, W., Yoshida, H., Koizumi, K., Yamada, H., Abe, K., Arimura, A., Yamane, N., Takahashi, K., Yamane, A., Oda, A., Tanaka, Y., Takemoto, H., Ohnishi, Y., Ikeda, Y., and Miyakawa, Y. (2008) The effect of a novel, small non-peptidyl molecule butyzamide on human thrombopoietin receptor and megakaryopoiesis, *Haematologica 93*, 1495–1504.
- 22. Nogami, W., Yoshida, H., Koizumi, K., Yamada, H., Abe, K., Arimura, A., Yamane, N., Takahashi, K., Tanaka, Y., Yamane, A., Takemoto, H., Ohnishi, Y., Ikeda, Y., and Miyakawa, Y. (2007) A novel, small non-peptidyl butyzamide activates human thrombopoietin receptor and promotes megakaryopoiesis, *Blood 110*, 654A–654A.
- Erickson-Miller, C. L., Delorme, E., Tian, S. S., Hopson, C. B., Landis, A. J., Valoret, E. I., Sellers, T. S., Rosen, J., Miller, S. G., Luengo, J. I., Duffy, K. J., and Jenkins, J. M. (2009) Preclinical activity of Eltrombopag (SB-497115), an oral, nonpeptide thrombopoietin receptor agonist, *Stem Cells 27*, 424–430.
- Delorme, E. O., Duffy, K. J., Lamb, P. I., Luengo, J. I., Tian, S. C. (2002) Regulated activation of cellmembrane receptors by metal-chelating agonists, Smithkline Beecham Corporation, Ligand Pharmaceuticals, *Intl. Patent Appl. No. PCT/US2001/* 050777, p 19.
- Kim, M. J., Park, S. H., Opella, S. J., Marsilje, T. H., Michellys, P. Y., Seidel, H. M., and Tian, S. S. (2007) NMR structural studies of interactions of a small, nonpeptidyl Tpo mimic with the thrombopoietin receptor extracellular juxtamembrane and transmembrane domains, *J. Biol. Chem.* 282, 14253– 14261.
- Palmiter, R. D. (2004) Protection against zinc toxicity by metallothionein and zinc transporter 1, *Proc. Natl. Acad. Sci. U.S.A.* 101, 4918–4923.
- Palmiter, R. D., and Findley, S. D. (1995) Cloning and functional-characterization of a mammalian zinc transporter that confers resistance to zinc, *EMBO J.* 14, 639–649.
- Michael, S. F., Kilfoil, V. J., Schmidt, M. H., Amann, B. T., and Berg, J. M. (1992) Metal-binding and folding properties of a minimalist Cys2His2 zinc finger peptide, *Proc. Natl. Acad. Sci. U.S.A.* 89, 4796– 4800.
- Nyborg, J. K., and Peersen, O. B. (2004) That zincing feeling: the effects of EDTA on the behaviour of zincbinding transcriptional regulators, *Biochem. J.* 381, e3-4.
- Frederickson, C. J., Suh, S. W., Koh, J. Y., Cha, Y. K., Thompson, R. B., LaBuda, C. J., Balaji, R. V., and Cuajungco, M. P. (2002) Depletion of intracellular zinc from neurons by use of an extracellular chelator in vivo and in vitro, *J. Histochem. Cytochem. 50*, 1659–1662.
- Tarasova, A., and Winkler, D. A. (2009) Modelling atypical small-molecule mimics of an important stem cell cytokine, thrombopoietin, *ChemMedChem* 4 2002–2011.

- Alper, P., Marsilje, T., Chatterjee, A., Lu, W., Mutnick, D., Roberts, M. J., He, Y. (2007) Heterotetracyclic compounds as TPO mimetics, Novartis Research Foundation, *Intl. Patent Application PCT/US2006/* 027691.
- Dexter, T. M., Garland, J., Scott, D., Scolnick, E., and Metcalf, D. (1980) Growth of factor-dependent hemopoietic precursor cell lines, *J. Exp. Med.* 152, 1036–1047.